Fully automatic identification and discrimination of sperm’s parts in microscopic images of stained human semen smear

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ABSTRACT

In the last years, digital image processing and analysis are used for computer assisted evaluation of semen quality with therapeutic goals or to estimate its fertility by means of spermatozoon motility and morphology. Sperm morphology is assessed routinely as part of standard laboratory analysis in the diagnosis of human male infertility. Nowadays assessments of sperm morphology are mostly done based on subjective criteria. In order to avoid subjectivity, numerous studies that incorporate image analysis techniques in the assessment of sperm morphology have been proposed. The primary step of all these methods is segmentation of sperm’s parts. In this paper, we have proposed a new method for segmentation of sperm’s Acrosome, Nucleus, Mid-piece and identification of sperm’s tail through some points which are placed on the sperm’s tail, accurately. These estimated points could be used to verify the morphological characteristics of sperm’s tail such as length, shape and etc. At first, sperm’s Acrosome Nucleus and Mid-piece are segmented through a method based on a Bayesian classifier which utilizes the entropy based expectation–maximization (EM) algorithm and Markov random field (MRF) model to obtain and upgrade the class conditional probability density function (CCPDF) and the apriori probability of each class. Then, a pixel at the end of sperm’s Mid-piece, is selected as an initial point. To find other pixels which are placed on the sperm’s tail, structural similarity index (SSIM) is used in an iterative scheme. In order to stop the algorithm automatically at the end of sperm’s tail, local entropy is estimated and used as a feature to determine if a point is located on the sperm’s tail or not. To compare the performance of the proposed approach with those of previous approaches including manual segmentation, the Accuracy, Sensitivity and Specificity were calculated.

1. INTRODUCTION

Infertility is a common clinical problem which causes considerable morbidity, including stress, depression and sexual dysfunction, in those couples affected [1]. The main cause of infertility is an anomaly of the sexual reproductive system. High percentage of these problems are from the male and finding ways to resolve this will be helpful to the physicians for a better and faster cure for couples. To determine infertility, some physica characteristics of the seminal plasma (such as smell, viscosity, pH and aspect) and spermatozoon’s parameters (such as concentration mobility and morphology) are analyzed [2]. Visual assessments of sperm by experts or CASA (Computer Aided Sperm Analysis) systems are the classical ways to determine the potential fertility of men. Manual methods are subjective and have led to widely varying results due to numerous factors such as different staining procedures, experience of technicians and human errors. So, manual procedures are inexact, subjective no repeatable and difficult to teach [3]. The disadvantage mentioned above makes difficult to interpret accurately the data and remarks the need of objective, precise, and repeatable techniques to study sperm. Due to the
complexity of sperm quality estimation, computerized techniques are essential tools. The majority of these
computer methods have been developed to analyze human sperm morphology and have afterwards been adapted
for other species [4]. The development of new methodologies is an ongoing research activity [5,6]. These
researches have enriched the available knowledge on sperm cells [7] and furthermore, digital image analysis has
allowed to classify subpopulations [8] or to describe shape abnormalities [6]. Most of these approaches use CASA
systems [9,10] that deploy image processing techniques or propose new description and classification methods
[11-14]. In this way, Sánchez et al. [4,15] proposed a technique to compute the fraction of boar spermatozoic
acrosome intact (class 1) or acrosome-damaged (class 2) in an optical phase-contrast microscope. As the
preprocessing step for segmentation of sperm's head, they used morphological closing, holes in the contours of the heads were filled and the spermatozoids tails were removed. In the next segmentation stage, spermatozoid heads were separated from the background deploying Otsu's method [16] to find a threshold that separates the heads from the background. Bieh et al. [17] applied Learning Vector Quantization (LVQ) in automated boar semen quality assessment. The classification of single boar sperm heads into healthy (normal) and non-normal ones was based on grey-scale microscopic images only. They used the same method proposed by Sánchez et al. [15] for segmentation of sperm's head. Alegre et al. [18] utilizing learning vector quantization (LVQ), suggested an automatic method to classify single sperm cells at acrosome intact (class 1) or acrosome-damaged (class 2) in an optical phase-contrast microscope. As the preprocessing step, Sperm head images were cropped manually from such a boar semen sample image. In each sperm head image they segmented automatically the sperm head by binarization using Otsu's method [16] and applying several morphological operations (dilations and erosions). Nowshiravan et al. [19] introduced a multisteps algorithm for sperm segmentation in microscopic image. At first the operator clicked on one chosen sperm their software defined a square that is bigger than the sperm, this square had both the tail object and the separation threshold. And form this they obtained a Histogram image, and imposed this on the threshold image and at the end all the objects were obtained with the same light surface. They used some image enhancement methods to improve their pictures, these steps removed sperm tail and middle part of it which was not clear earlier, but show the head of it better than before. Nafisi et al. [20] proposed a segmentation algorithm based on $\epsilon$ threshold level for finding sperms in low contrast images. First, an image enhancement algorithm was applied to remove extra particles from the image. Then, the foreground particles (including sperms and round cells) were segmented form the background. Finally, based on certain features and criteria, sperms were separated from other cells. Park et al. [21] proposed a method based on the Hough transform for the quantitative estimation of the morphological characteristics of the sperm. Images of the sperms were acquired into the digital format using the optical microscope, CCD camera, and flame grabber. For each sperm in the image, the region of interest for the segmentation of the sperm head was selected using the density difference between the sperm head and the background. The boundary of the sperm head was approximated with an ellipse and was used for estimation of the morphological characteristics of the sperm. Carrillo et al. [22,23] introduced an approach called nth-fusion for segmentation of sperms Acrosome, Nucleus and Midpiece in a computer aided tool for the objective analysis of human sperm morphology, commonly known as Automated Sperm Morphology Analyzer (ASMA). After enclosing individual sperms (head and mid-piece) using bounding boxes, they used nth-fusion method which was based on nth-level thresholding of an image followed by intersection with n special masks. In order to obtain the desired segmentation results, aprior objects morphological model, which was based on the information fusion technique, was used. Finally, based on certain features and criteria, sperms were separated from other cells. Abbiramy et al. [24] performed a method for segmenting objects in microscopic images into its constituent's parts based on morphological operators and edge detections.

In this paper we have proposed a fully automatic method for identification and discrimination of sperm's
Acrosome, Nucleus, Mid-piece and tail that requires no training and atlas. The proposed method includes two
major modules:

• Segmentation of sperm’s Acrosome, Nucleus and Midpiece;

• Localized identification and discrimination of sperm’s tail.

At first, an improved hybrid method [25] is used to remove noise from the sperm image (R component of RGB
color image). Then, a simple threshold is applied to build a primary mask containing sperm’s Acrosome, Nucleus
Mid-piece, and also small objects in seminal plasma. The small objects have been eliminated and to build the final
localized mask, the minimum area bounding box of each individual region is computed through the Rotating
Callipers method [26,27]. The detection rate and speed has been increased using the bounding boxes. Pixels
inside the bounding boxes are considered as samples. Then, intensities of samples are modeled using a gaussian
mixture model which consists of three kernels as Background, Nucleus, and a class of Acrosome and Midpiece
This step starts with only one kernel and uses an entropy based EM algorithm to estimate three kernels as three
mentioned classes in an automatic manner which does not need initial values for parameter estimation. After
estimation of the model, we would be able to classify brain pixels by knowing apriori probabilities of the classes. The next step is obtaining these apriori probabilities with no training and atlas. So, a MRF model and EM algorithm are applied to update and attain apriori probabilities and means and variances of each class. Finally, Samples in bounding boxes are classified using Bayesian classification [28,29].

After localized segmentation of sperm’s Acrosome, Nucleus and Mid-piece, the pixel at the distal point of sperm’s Mid-piece is considered as an initial point. The proposed method uses a structural similarity index [30] and Rényi entropy [31] in an iterative scheme to estimate sperm’s tail with some points which are placed on the sperm’s tail accurately [32]. These estimated points can be used to analyze characteristics of sperm’s tail such as length shape, and etc. In the next sections, details of the research procedure including segmentation and identification of sperm’s Acrosome, Nucleus, Mid-piece and tail are explained. Early and encouraging experiments with these methods have been presented in [32].

2. MATERIALS AND METHODS

2.1. Image Acquisition Technique

Sample images were acquired from modified Papanicolaou stained sperm smears. Fresh sperm samples were incubated for 30 to 60 minute in 37˚ Celsius. The smear was then prepared after complete liquefaction and the slides were dried in the air before staining with modified Papanicolaou method. The images were captured by means of a 560 TV-line CCD camera mounted on the third eyepiece of a trinocular direct microscope (Proway BK5000) with a total magnification of 1000× using Plan Achromatic Infinity objective lenses and a resolution of 576×764 pixels in RGB color space. 10 to 25 Images of different fields were captured from each slide. And totally 100 slides were analyzed (each slide consists of 1 to 5 sperms).

2.2. Preprocessing

To create a primary mask containing sperm’s Acrosome, Nucleus and Mid-piece, the Red component of RGB color image is used. The Red component contains most of the information associated with the darkest color, which domains the head. The images first had to be scaled. Therefore the range between zero intensity and maximum intensity, M, in the original 12-byte data (I) was scaled to a new intensity (I\textsubscript{s}) between 0 and 255 (8-bit) which is obtained by I\textsubscript{s} = I/M × 255.

2.2.1. Remove Noise from R Component

To remove noise, an improved hybrid method [25] is applied to sperm image (R component of RGB color image). This method consists of two stages. The first stage consists of a fourth order partial differential equation (PDE) and the second stage is a relaxed median filter, which processes the output of fourth order PDE. This model enjoys the benefit of both nonlinear fourth order PDE and relaxed median filter. By using a relaxed median filter we can preserve more image details than the standard median filter. This method preserves fine details, sharp corners and thin lines and curved structures to a large extent. The L2-curvature gradient flow method of You et al [33] is used in this model:

\begin{equation}
\frac{\partial u}{\partial t} = \lambda \nabla^2 u - \lambda \left( \lambda \left| \nabla u \right|^2 \nabla u \right)
\end{equation}

where \(\Delta u\) is the Laplacian of the image u. Since the Laplacian of an image at a pixel is zero if the image is planar in its neighborhood, the PDE attempt to remove noise and preserve edges by approximating an observed image with a piecewise planar image. The desirable diffusion coefficient \(\lambda\) should be such that (1) diffuses more in smooth areas and less around less intensity transitions, so that small variations in image intensity such as noise and unwanted texture are smoothed and edges are preserved. Another objective for the selection of \(\lambda\) is to incur backward diffusion around intensity transitions so that edges are sharpened, and to assure forward diffusion in smooth areas for noise removal. The PeronaMalik diffusivity function [34] is used in the implementation as below:

\begin{equation}
\lambda(x) = \frac{1}{1 + \left( \frac{\nabla u}{\nabla u(x)} \right)^2}
\end{equation}

The discrete form of non-linear fourth order PDE described in Eq.1 is as follows:
where

\[ \Delta t \] is the time step size and \( h \) is the space grid size. Relaxed median filter [35,36] is used in combination with Eq.1 to remove large spike noises. The proposed hybrid method by Rajan et al. [25] is defined as follows:

\[ \text{RM} = \text{relaxed median filter with lower bound } \underline{B} \text{ and upper bound } \overline{B}. \]

If \( \text{filtered} \) is the output of a relaxed median filter, then \( \text{filtered} \) can be written as

\[ \text{filtered} = \text{median value of the samples inside the window}. \]

The sliding window \( \text{window} \) is to be the window located at position \( i \). The lower bound and upper bounds for relaxed median used in the experiments are 3 and 5 respectively. By using a relaxed median filter we can preserve more image details than the standard median filter. This method preserves fine details, sharp corners and thin lines and curved structures to large extent. Then, a simple threshold is applied to build a primary mask, \( \text{primary mask} \), which contains sperm's Acrosome, Nucleus and Mid-piece, and also small objects in seminal plasma. The threshold value is calculated according to

\[ \text{threshold value} = \mu - k \sigma, \]

in which \( \mu \) and \( \sigma \) are mean and standard deviation of the noise-removed image, respectively. The primary mask, \( \text{primary mask} \) is defined as

\[ \text{primary mask} = \text{primary mask}. \]

The small objects have been eliminated and this mask, containing sperm's Acrosome, Nucleus and Mid-piece is used to build the final mask at the next step. It is reminded that \( \text{threshold value} \) is not enough accurate to detect sperm's Acrosome, Nucleus, Mid-piece areas in all sperm images (especially about the distal points of Mid-piece). So, \( \text{primary mask} \) has been only used as a primary mask to build the final mask containing all candidate pixels which may belong to the sperm's head and Mid-piece.

### 2.2.2. Finding the Best-Fitted Rectangle for Each Region

To build the final mask, the minimum area bounding box of each individual region is computed through the Rotating Calipers method [26,27]. This method is capable of computing the minimum area enclosing rectangle in linear time. To apply Rotating Calipers method, the two dimensional convex hull of all visible points (for each region) is computed using the monotone chain algorithm [37]. This algorithm is linear with respect to the number of input points \( O(n) \), assuming that input points are sorted by increasing \( x \) and increasing \( y \) coordinates. The minimum rectangle enclosing a convex polygon \( P \) has at least one side collinear to one edge of \( P \) [26], using this property, a brute-force approach would be to construct an enclosing rectangle for each edge of \( P \). This has \( \epsilon \) complexity of \( O(n^2) \) since we have to find minima and maxima for each edge separately. The rotating calipers algorithm rotates two sets of parallel lines (calipers) around the polygon and incrementally updates the extreme values, thus requiring only linear time to find the optimal bounding rectangle. **Figure 1** [38] illustrates one step of this algorithm: The support lines are rotated (clock-wise) until a line coincides with an edge of \( P \). If the area of the new bounding rectangle is less than the stored minimum area rectangle, this bounding rectangle becomes the new minimum. This procedure is repeated until the accumulated rotation angle is greater than 90 degrees.
Figure 2 shows the obtained results for each step

**Figure 1.** Rotating calipers.

**Figure 2.** Result of applying an improved hybrid method and Rotating Calipers algorithm to a typical sperm image: (a) The original R component of RGB color image; (b) Noise removed image; (c) Selected objects after applying threshold; (d) Primary mask without small objects; (e) Final mask which is created using Rotating Calipers method; (f) Sperm image overlaid on its bounding boxes.

when the proposed method is applied to a typical sperm image. The proposed method for creating a mask containing sperm’s Acrosome, Nucleus and Mid-piece is summarized as below:

- Selection of the R component of RGB color image.
- Removing noise from R component using an improved hybrid method.
- Building a primary mask containing sperm’s Acrosome, Nucleus, Mid-piece, and also small objects in seminal plasma, by applying a simple threshold to the noise removed image.
- Elimination of small objects.
- Finding the best-fitted rectangle for each region using rotating calipers.

### 2.3. Problem Formulation

Support that $X$ is a $n$-dimensional random variable and comes from a Gaussian mixture model of $M > 1$ components. Then the probability density function of the Gaussian mixture model can be repressed as the following:

$$f(x) = \sum_{m=1}^{M} \pi_m \phi(x; \mu_m, \Sigma_m)$$

where $\pi_m$ corresponds to the weight of each component which satisfies $\sum_{m=1}^{M} \pi_m = 1$. For the Gaussian mixtures model, each component density $\phi(x; \mu_m, \Sigma_m)$ is a Gaussian probability density given by:

$$\phi(x; \mu_m, \Sigma_m) = \frac{1}{(2\pi)^{n/2} |\Sigma_m|^{1/2}} \exp\left(-\frac{1}{2} (x - \mu_m)^T \Sigma_m^{-1} (x - \mu_m) \right)$$

where $T$ denotes the transpose operation, $\mu_m$ is the mean vector and $\Sigma_m$ is the covariance matrix which is assumed positive definite. Here we encapsulate these parameters into a parameter vector, writing the parameters of each component as $\theta_m$, to get

$$\theta = \{ \theta_1, \theta_2, \ldots, \theta_M \}$$

Eq.11 can be rewritten as:

$$f(x) = \sum_{m=1}^{M} \pi_m \phi(x; \mu_m, \Sigma_m)$$

If we knew the component from which $x$ came, then it would be simple to determine the parameters. Similarly if we knew the parameters, we could determine the component that would be most likely to have produced $x$. The difficulty is that we know neither.

### 2.4. Bayesian Classification

Bayesian Classification is a probabilistic technique of pattern recognition and is based on the principle of Bayes decision theory \[39\], given in Eq.14 below...
where, $x$ is a given feature vector, $\Theta$ denotes a class, or state of nature, $\pi_\Theta$ is the prior probability of class $\Theta$, $\pi_x$ is prior probability of the feature vector $x$, $\pi_{\Theta|x}$ is aposteriori probability, which a feature vector should be classified as belonging to class $\Theta$, $p(x|\Theta)$ is the conditional probability that a feature vector occurs in a given class $\Theta$. For the approach here, the feature $x$ shall consist of one component, intensity of brain pixels. The quantity $\tau$ is known as the evidence, and serves only as a scale factor, such that the quantity in Eq.14 is indeed a true probability, with values between zero and one. So, the maximum a posteriori (MAP) estimate of Eq.14 is used as below

\[ \text{Eq.15} \]

According to Bayesian theory [40], the feature vector $x$ is classified to $\Theta$ of which the aposteriori probability given $x$ is the largest between the classes.

\[ \text{Eq.16} \]

Bayes decision rule is optimal in the sense of minimization of the probability of error. It is quite obvious that such an Ideal Bayesian solution can be used only if distributions $\pi_\Theta$, and the apriori probabilities $\pi_x$ are known. In the context of classification of brain tissue, the probability models are not known, and therefore must be approximated. The performance of the Bayesian classifier is directly related to how well these distributions can be modeled.

### 2.5. Segmentation of Sperm’s Acrosome, Nucleus and Mid-Piece

Sperm’s Acrosome, Nucleus and Mid-piece are segmented using a fully automatic method which is based on entropy based EM algorithm and Markov random field model [29]. This method estimates a gaussian mixture model with three kernels as Background, Nucleus and a class of Acrosome and Mid-piece. To estimate this model, an automatic Entropy based EM algorithm [28] was used to find the best estimated Model. Then, Markov random field (MRF) model and EM algorithm were utilized to obtain and upgrade the class conditional probability density function and the apriori probability of each class. After estimation of Model parameters and apriori probability, samples in bounding boxes were classified using Bayesian classification.

Based on the explanations mentioned above, the block diagram of our method for segmentation of sperm’s Acrosome, Nucleus and Mid-piece is shown in Figure 3.

**Figure 3.** Block diagram of the proposed approach for fully automatic segmentation of sperm’s Acrosome, Nucleus and Midpiece.

and is summarized below

1) Smoothing the R component of RGB color image using a Gaussian filter.

2) Selection of pixels inside the bounding boxes as input samples.

3) Estimation of input samples distribution using Entropy based EM algorithm and Markov Random Field Model [29] with three kernels as Background, Nucleus and a class of Acrosome and Mid-piece.

4) Bayesian classification of the input image (only pixels inside the bounding boxes) using the obtained Gaussian Mixture Model.

5) Postprocessing: Acrosome and Mid-piece are classified into the same class (i.e., two separated areas in one class). These two areas are grouped into two separated classes (i.e., Acrosome and Mid-piece) using their positions respect to the Nucleus and bounding box corners. At first, the distance between each corner of bounding box and center of Nucleus is computed. Then, the corner whose value is less than others is considered as origin. Acrosome is the region whose distance from origin is less than Mid-piece (other region).

### 2.6. Simulation Results: Segmentation of Sperm’s Acrosome, Nucleus and Mid-Piece
Figure 4 shows the results of the proposed algorithm for a typical sperm image, including estimated distribution obtained through the Entropy based EM algorithm and MRF model overlaid on distribution of the samples (inside the bounding boxes). In Figure 5, results of applying the proposed algorithm to other sperm images have been shown.

2.7. Identification of Sperm’s Tail

After localized segmentation of sperm’s Acrosome, Nucleus and Mid-piece, the pixel at the distal point of sperm’s Mid-piece is considered as an initial point. The method [32] uses a structural similarity index [30] and Rényi entropy [31] in an iterative scheme to estimate sperm’s tail with some points which are placed on the sperm’s tail accurately. These estimated points can be used to analyze characteristics of sperm’s tail such as length, shape and etc.

Let X be a digital image of size (M × N) which consists of a sperm, and A be a pixel on the sperm’s tail which is considered as an initial point (point A). Figure 6(a) shows a typical sperm image, a section of sperm’s tail it zoomed in and shown in Figure 6(b). Suppose the bright green pixel is point A, to find the next pixel which is on the sperm’s tail and has the highest structural similarity index (SSIM) with respect to point A, we have used two widows (w₁: p × p) and w₂: k × k, k < p).

w₁ is centered at point A and is applied to limit A’s neighborhood. Figure 6(b) shows w₁ in pink. w₂ is a sliding window which moves over the boundaries of w₁ and computes the local structural similarity (SSIM) index for each pixel with respect to point A. Figure 6(c) shows pixels which are placed on the boundaries of w₁ in deep pink, also different positions of the sliding window (w₂) which moves over the boundaries of w₁ is shown in blue. So, the SSIM index is calculated within the sliding window (w₂) for all pixels which are placed on the boundaries of w₁ to find the pixel with the highest SSIM index. SSIM index for all pixels are calculated and scaled ([0,255]), these values are shown in Figure 6(d). The pixel (Point B) which has the highest SSIM index and pixel A are shown in Figure 6(e). If we do the same algorithm for new selected pixel (B), it is possible to select previous pixel (A) as a new pixel. So, before running the new iteration for the new selected pixel, a neighborhood of previous pixel is changed to a different value (say zero). This process is done through another window (w₃: l × l, k < l < p). Figure 6(f) shows this process.

This algorithm is implemented in an iterative scheme, and for every new selected pixel, local entropy is computed to detect if we have reached the end of the tail or not.

In this way, for every new selected pixel, the local entropy is estimated. To estimate the local entropy for every new selected pixel, a window with odd size (w₄: h × h) is centered at that pixel, then for every pixel which is placed inside the window, only Red and Green components are extracted as a two dimensional feature. The local entropy is estimated by estimation of the entropy of these samples. Figure 7 shows variations of the local
interpretation of the references to color in this figure, the reader is referred to the web version of this article).

Figure 7. Result of applying the proposed algorithm to the sperm’s image: (a) Estimated points, as the result of proposed method; (b) Variations of the local entropy for estimated points on the sperm’s tail; (c) The mean value of local entropies, for every five selected pixels.

Entropy on the sperm’s tail. As it is seen, by reaching the end of sperm’s tail, the local entropy decreases. So, by finding a fitted threshold we would be able to stop the algorithm at the end of sperm’s tail, automatically. We have experimentally observed that a two dimensional feature which contains Red and Green components, gives more strict threshold in comparison with other features.

For a fitted threshold, if the estimated value is greater than the threshold, it means that the selected pixel is placed on the sperm’s tail. In other words, the local entropy is considered as a criterion which shows that if we are on the sperm’s tail or not. Finding such a threshold for all sperms is a challenging task. To tackle this problem, the localized thresholding is used. For every five new selected pixels, the local entropy is computed. Then, the mean value of local entropies is considered to compare with a threshold which is called window-threshold. The window-threshold is a fitted threshold for all sperms, and is used to find out if we are close to the distal points of sperm’s tail or not. So, for every five new selected pixels, local entropies are computed and mean value is compared with the window-threshold, if it is greater than the window-threshold, it means that we are not close to the distal points of sperm’s tail and the algorithm must be continued to find five new estimated points in the next five iterations. Otherwise, if the mean value is less than window-threshold, we are close to the distal points and the algorithm must be continued by individual thresholding for every new selected pixel to stop the algorithm in the next five iterations which will be the last ones. Means that from this moment onwards, the algorithm will find at most five new estimated points, in next iterations. Because, by checking the window-threshold, we have found out that we are close to the distal points of sperm’s tail. For individual thresholding, the minimum local entropy (MLE) of five previous selected pixels is used for thresholding as below

\( \text{MLE} \) has been experimentally set to 0.9 for the best result.

The algorithm for detection of sperm’s tail includes two major modules:

- Selecting an initial pixel (A) which is placed at the end of mid-piece, and finding the pixel (B) with the highest SSIM index respect to that. To compute and compare the structural similarity (SSIM) index, only the Red component of the RGB color image is considered as a separate image and is used to perform this module.

- Verifying if the selected pixel (B) is on the sperm’s tail or not, this procedure is based on the local entropy estimation. To evaluate the local entropy, only the Red and Green components of RGB color image are used as two dimensional feature.

The steps of proposed method are described in detail as below:

1) Setting pixels on the sperm’s head and mid-piece to a different value (say zero).

2) Smoothing the input image using an average filter.

3) Selecting the distal point of sperm’s Mid-piece as the initial point, pixel (A), and setting the Counter to one.

4) Centering the window \( w_1 \) at pixel A.

5) Moving a sliding window \( w_2 \) over \( w_1 \)’s boundaries and compute SSIM index, locally, through Eq.12, for each pixel over the boundaries of \( w_1 \) and pixel A.

6) Finding the pixel with the largest amount of SSIM Index (B).

7) Estimation of the local Entropy at pixel B.

8) Setting pixels around the new selected pixel (B) to a different value (say zero), through the window \( w_3 \).

9) Replacing the initial pixel (A) with the new selected pixel (B).
10) Checking the Counter: if the Counter is equal to five, go to the next step, otherwise increase the Counter by one and go to the Step 4.

11) Checking the localized thresholding: compute the mean value of the local entropies of the last five selected pixels. If it is greater than the window-threshold, set the Counter to one and go to the Step 4, otherwise find the minimum local entropy (MLE) of the last five selected pixels and go to the next step.

12) Repeating Steps 4-9 to find the new estimated pixel.

13) Comparing the local entropy of the new selected pixel with the threshold (Eq.17). If it is greater than the threshold, go to the step 12 to find a new pixel on the sperm’s tail, otherwise stop the algorithm.

2.8. Simulation Results: Identification and Discrimination of Sperm’s Tail

The results of the proposed method, for different sperms are shown in Figure 8. Figures 8(a)-(d) show the original sperm images. The proposed method is applied to sperm images, and the estimated points are shown in Figures 8(e)-(h). As it is seen, the estimated points are exactly located on the sperm’s tail. Figures 8(i)-(l) show how the estimated local entropy changes with the position of estimated points on the sperm’s tail. The last points in each tail have the lowest value of local entropy in comparison with the other points, these values are less than the threshold, and the algorithm was stopped because of these points.

3. EVALUATION

The algorithms were tested with a 100 slides database with multiple spermatozoa (including 283 sperms). The first stage tested was the “detection and extraction of individual spermatozoon” with the entire image database. In this research, criteria Accuracy ($A_c$), Sensitivity ($S_e$) and Specificity ($S_p$) are used for evaluation.

\[
A_c = \frac{TP}{TP + FP} \\
S_e = \frac{TP}{TP + FN} \\
S_p = \frac{TN}{TN + FP}
\]

In these equations, TP is the number of true positive, FP is the number of false, TN is the number of true negative, and FN is the number of false negative. The evaluation is summarized in Table 1.

Also the, Accuracy of sperm’s head, Acrosome, Nucleus and Mid-piece are computed 94.3%, 92.4%, 95.1% and 90.2%, respectively. For all sperms in database, sperm’s tail length is computed manually and compared with the estimated value through the estimated points. Success rate [32] is defined below

\[
\text{Success Rate} = \frac{\text{Number of Correctly Estimated Tail Lengths}}{\text{Total Number of Sperms}}
\]

and computed for all sperms. The overall Success Rate of this approach is 97.6%.

4. DISCUSSION

In this paper a new approach, for fully automatic identification and discrimination of sperm’s Acrosome, Nucleus Mid-piece and tail in microscopic images of stained human semen smear that requires no training and atlas, is proposed. The proposed method includes two major modules. At first, to increase the detection rate and speed, a localized mask containing sperm’s Acrosome, Nucleus and Mid-piece is built through the minimum area bounding boxes. This procedure is done using an improved hybrid method and the Rotating Calipers algorithm. Pixels inside the bounding boxes are considered as samples. Distribution function of samples is estimated by

Figure 8. Result of applying the proposed algorithm to different sperms images: (a)-(d) Original sperm images; (e)-(h) Estimated points, as the result of the proposed method; (i)-(l) Use of local entropy changes as a measure for identification of end of tails.
Table 1. Evaluation results for detection and extraction of individual spermatozoon.

<table>
<thead>
<tr>
<th>Method</th>
<th>Accuracy (A_c)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Proposed Method</td>
<td>0.95</td>
</tr>
<tr>
<td>Carrillo et al. [22]</td>
<td>0.88</td>
</tr>
</tbody>
</table>

As it is seen, pixels which are placed on the sperm’s tail have higher Luminance comparison components in comparison with others. Also, in addition to pixels which are placed on the sperm’s tail, there are some other pixels over the boundary which have higher values for contrast and Structure comparison components. So Luminance comparison component has the most relative importance.

As mentioned before, structural similarity index satisfies the Boundedness condition (SSIM < 1), that’s why α, β and γ have been experimentally set to 0.1, 0.5 and 0.9 for the best result.

One of the important advantages of the proposed method is the ability to detect sperm’s tail in the low-contrast sections. Also, the execution time of implemented algorithm is too low. Our proposed approach is evaluated via Accuracy, Sensitivity, Specificity and Success Rate in a data set of microscopic images of stained human semen smear.

Some other methods have been developed to analyze human sperm morphology, the primary steps of all these
methods is sperm segmentation and discrimination. Sánchez et al. [4,15] and Nowshiravan et al. [19] propose methods based on morphological operators and thresholding to segment sperm’s head. Because the Acrosome and the Mid-piece have similar characteristics (intensity level, texture) the segmentation with traditional techniques (thresholding, region growing) does not give good results [19,20]. To tackle this problem, Carrillo et al. [22, 23] presented an approach called nth-fusion for segmentation of sperm’s Acrosome, Nucleus and Mid-piece in a computer aided tool for the objective analysis of human sperm morphology, commonly known as Automatic Sperm Morphology Analyzer (ASMA). After enclosing individual sperms (head and Mid-piece) by bounding boxes, they used nth-fusion method which was based on nth-level thresholding of an image followed by intersection with n special masks. In order to obtain the desired segmentation results, aprior objects morphological model, which was based on the information fusion technique in a feature level was used. For each segmented sperm in image, they had to run the algorithm to detect sperm’s parts. It is reminded that Carrillo et al. [22] use manual segmentation for evaluation of their methods. We, too, used manual segmentation for evaluation. They used similar methods of evaluation. Therefore comparison of our method with this method is reasonable. This comparison is done in Table 2. As it is seen in Table 2, the proposed method in this paper improves the accuracy of segmentations. The drawback of the proposed method for detection and discrimination of sperm’s tail is that objects which are placed on the sperm’s tail, affect the performance of the proposed method. So, in future investigations we intend to use various geometric features to increase accuracy values.

REFERENCES


Semen analysis comprises a set of descriptive measurements of spermatozoa and seminal fluid parameters that help to estimate semen quality (4). Conventional semen analysis includes measurement of particular aspects of spermatozoa such as concentration, motility and morphology and of seminal plasma. Quantification and identification of non-spermatozoidal cells and detection of antisperm antibodies are also part of basic semen analysis (6). Normal values of semen parameters issued by the World Health Organisation (WHO) in 1992 are generally used as reference values (Table I). The Papanicolaou smear for staining of spermatozoa is the method most widely used in andrology laboratories. In each sperm head image they segmented automatically the sperm head by binarization using Otsu's method [16] and applying several morphological operations (dilations and erosions). Nowshiravan et al. [19] introduced a multi steps algorithm for sperm segmentation in microscopic image. Sample Images were acquired from modified Papanicolaou stained sperm smears. Fresh Sperm samples were incubated for 30 to 60 minute in 37°C Celsius. Sperm's Acrosome, Nucleus and Mid-piece are segmented using a fully automatic method which is based on entropy based EM algorithm and Markov random field model [29]. This method estimates a gaussian mixture model with three kernels as Background, Nucleus and a class of Acrosome and Mid-piece. Samples were stratified based on total sperm count, spermatozoa concentration, progressive motility, total motility and spermatozoa morphology. Microbiota profiling was performed using 16S rRNA gene amplicons sequencing and total bacterial load was determined using a panbacterial quantitative PCR. Our study indicates that overall bacterial content of sperm might not play a major role in male infertility. Although no major shifts in microbiota composition or diversity were found, the differential abundance of specific bacterial genera in the sperm suggests that a small... WHO Laboratory Manual for the Examination and Processing of Human Semen, 5th Edn. Geneva: WHO Press. Google Scholar.